



Mesenchymal Stem Cells As A Promised Therapy For Alzheimer's Disease

Kornpaka Suntipong

Mahidol University International Demonstration School, Salaya, Phutthamonthon,
Nakhon Pathom, Thailand 73170

***Corresponding Author:**

Kornpaka Suntipong

Mahidol University International Demonstration School, Salaya, Phutthamonthon,
Nakhon Pathom, Thailand 73170

Type of Publication: Original Research Paper

Conflicts of Interest: Nil

Abstract

Alzheimer's Disease (AD) is a chronic neurodegenerative condition and has been classified as the most prevalent classification of dementia in the present day. It manifests itself in the form of an abnormal build-up of protein in the medial entorhinal cortex and the hippocampus, leading to cognitive malfunction. In the present day, there are no known cures for AD; instead, the condition is treated with prescription drugs to help relieve the symptoms. However, recent studies on mesenchymal stem cell transplantations have found that promising results such as improvement of spatial memory, decreased levels of pro-inflammatory cytokines, and reduction in amyloid-protein plaques as well as phosphorylated Tau protein. This article aims to introduce AD and its etiological hallmarks, to illustrate an overview of recent developments on mesenchymal stem cell research specific to AD, and to discuss the future prospects and potential of MSC-based therapies.

Keywords: Alzheimer's Disease, Mesenchymal stem cells, Stem cells

Introduction

Alzheimer's Disease (AD) is a cumulative neurodegenerative disease; classified as the most prevalent classification of dementia (Bondi, Edmonds, & Salmon, 2017). This neurodegenerative disease is caused by the progressive build-up of protein which leads to neuronal death and their connections in the regions of the brain concerned with cognitive function (Weller & Budson, 2018). Tau proteins are natively unfolded microtubule-associated proteins whose function depends on their phosphorylation state (Chong, Ng, Koh, & Chye, 2018; Nam, Lee, Moon, & Chang, 2020; Nu et al., 2018). Aggregated phosphorylation causes the accumulation of the tau protein in the somatodendritic neuron compartment which consequently forms Neurofibrillary tangles (NFTs) leading to neurons becoming unable to maintain their cytoskeleton (Gant et al., 2018; Pascoal et al., 2018; Pascoal et al., 2020). Amyloid plaques (also known

as senile plaques) are a pathological hallmark in the progression of AD (Ji et al., 2018; Zhan, Stamova, & Sharp, 2018). Accumulation of amyloid-beta ($A\beta$) has been implicated in the etiology of AD and is believed to be a characteristic factor in the neural damage in the regions of the hippocampus and cortex (Bayer & Wirths, 2010; Jiang et al., 2018; Martins et al., 2019; Song & Choi, 2013).

As of this time, there are no curative treatments for AD. However, acetylcholinesterase (AChE) inhibitors such as donepezil, galantamine, and rivastigmine are standard prescriptions to inhibit the breakdown of ACh by the enzyme cholinesterase; this helps to temporarily alleviate some symptoms of AD (Long & Holtzman, 2019; Mathew et al., 2019; Murray, Faraoni, Castro, Alza, & Cavallaro, 2013). Even so, these standard treatments cease to reach the desired clinical outcome, therefore leading to a surge of innovation for new treatments to improve therapeutic results for AD. Of many clinical trials and

treatments for a range of diseases, stem cell therapy has risen as an alternative regimen (Connors, Ames, Woodward, & Brodaty, 2018; Hanfelt, Peng, Goldstein, & Lah, 2018; Magalhães, Alves, Fortuna, Llerena, & Falcão, 2020). Recent studies, adipose-derived mesenchymal stem cells (MSCs) have proved to activate microglia, causing a reduction in the level of A β . This shows substantial promise in MSC-based treatments for AD (Chang, Kim, Joo, Ha, & Suh, 2014; Deng et al., 2019; Ma et al., 2013; Shigematsu, Takeda, Komori, Tahara, & Yamagishi, 2021; Yan et al., 2014). This paper aims to gather the molecular mechanism of MSC-based treatment of AD and the clinical outcomes in the use of MSC-based remedies for AD in clinical trials in order to broaden ideas of future treatment and to strengthen their potential for success.

The etiological mechanisms of AD and standard treatment

The first genetic mutations which were found to be linked to early-onset AD were mutations in the amyloid precursor protein (APP). APP is the precursor molecule that is cleaved proteolytically by the cleavage enzymes such as β -secretases and γ -secretases to biosynthesise 37 to 49 amino acid residue peptides, A β (Pluta, Furmaga-Jabłońska, Maciejewski, Ułamek-Kozioł, & Jabłoński, 2013; Schauenburg et al., 2018; Wang & Pei, 2018). These amino acids have proven to be the root source to the extracellular senile plaques found in patients with AD. Conventionally, APP is metabolized by two definite pathways: (1) the secretory pathway, and (2) the amyloidogenic pathway. In the secretory pathway (1), APP is cleaved by β -secretase which releases an N-terminal fragment (sAPP β) along with a C-terminal fragment (C99) which undergoes additional division by γ -secretases, resulting in the yield of monomers; A β peptides (Esmieu, Ferrand, Borghesani, & Hureau, 2021; Rana & Sharma, 2019). The monomers, in turn, progressively aggregates to dimmers, trimers, oligomers, protofibrils, and fibrils, leading to protein deposits (amyloid plaques). Soluble A β oligomers interact with neuronal and glial cells which cause various cellular dysfunction: induced cell death and neuronal apoptosis, impaired intracellular signalling pathways, and increased Tau protein phosphorylation (Fontana et al., 2020; Y. S. Kim, Jung, & Yoon, 2018; S. Li & Selkoe, 2020). A β fibrils form deposits of protein which are the

foundation of neuritic plaques in the regions of the cerebral cortex (Fontana et al., 2020; Y. S. Kim et al., 2018; S. Li & Selkoe, 2020). These plaques are deposited in progressive and sequential morphological forms throughout the progressive stages of AD (Fontana et al., 2020; Y. S. Kim et al., 2018; S. Li & Selkoe, 2020). The plaques cause the stimulation of astrocytes and microglia, damage to neuronal cells which cause the loss of synapses, and eventually, cell death (Azevedo et al., 2013; Bessis, Béchade, Bernard, & Roumier, 2007; Fontana et al., 2020; Y. S. Kim et al., 2018; S. Li & Selkoe, 2020).

Tau proteins are microtubule-associated neuronal phosphoproteins that are highly involved in the construction and stabilization of both the neuronal cytoskeleton and its microtubules, as well as the regulation of the axonal transport within neuronal axons. In neuronal axons, the microtubules are uniformly oriented due to the Tau protein (Aanandhi, Niventhi, Rujaswini, Hemalatha, & Praveen, 2018; Baas & Qiang, 2019; Qiang et al., 2018; Samra et al., 2017). During the process of remodeling the cell's cytoskeleton, changes in the state of phosphorylation occur; this makes the regulatory functions and mechanisms of the phosphoprotein essential to increase synaptic plasticity (Alavi Naini & Soussi-Yanicostas, 2015; Feng et al., 2020; Kimura et al., 2014). If the Tau protein were to go under abnormal or aggregated phosphorylation, this would affect its ability to bind to tubulin resulting in an agitated microtubule structure. Hyperphosphorylated Tau protein damages axonal transport and synaptic metabolism which brings about a loss of cell viability, causing the disintegration of microtubular cytoskeleton structures and neurone cell death (Boonen, van Tijn, & Zivkovic, 2009; Narayanan et al., 2020).

The interchange between Tau proteins and tubulin is imperative in the stabilization of the microtubular structure (Bossenmeyer-Pourié et al., 2019). The process consists of the promotion of polymerization by Tau protein which causes the inhibition of the fast depolymerization of tubulin; this is controlled by the balance of phosphorylation and dephosphorylation at the phosphorylation sites of the phosphoepitopes of Tau protein: serine and threonine. Both the phosphorylation and dephosphorylation of epitopes helps to aid the ability of Tau protein to interrelate with α - and β -tubulin (Bhandare, Kumbhar, &

Kunwar, 2019; Bossenmeyer-Pourie et al., 2019; Kellogg et al., 2018; Nasu, Furukawa, Suzuki, Takeuchi, & Koriyama, 2020).

Conventionally, a majority of serine and threonine phosphoepitopes in both fetal and Paired Helical Filament Tau (PHF-Tau) is accompanied by proline residues, raising the conclusion that Tau kinases are proline-directed kinases; enzymes that possess the ability to phosphorylate Tau protein. Such enzymes have been discerned in the brains of AD patients and in NFTs (Drepper et al., 2020; Hanger, Betts, Loviny, Blackstock, & Anderton, 1998; Hefti et al., 2019; Morris et al., 2021; Vincent, Zheng, Dickson, Kress, & Davies, 1998). This hyperphosphorylation of Tau protein by the proline-directed kinases causes abnormalities that impair its ability to bind to tubulin; causing an unstable microtubule structure (Hefti et al., 2019). Additionally, the aggregated deposition of such proteins also causes the deterioration of axonal transport, synaptic metabolism, cytoskeleton collapse, and neuronal death (Hefti et al., 2019; Vincent et al., 1998).

As of this time, there are no curative treatments for AD. However, AChE inhibitors such as tacrine, donepezil, galantamine, and rivastigmine were approved for inhibiting the breakdown of ACh by the enzyme cholinesterase. These drugs temporarily help to alleviate some symptoms of AD; though they are not curative treatments (Briggs, Kennelly, & O'Neill, 2016). An alternative to AChE inhibitors for those who do not respond or are allergic to AChE inhibitors is memantine (Murray et al., 2013). Memantine is an N-methyl D-aspartate (NMDA) antagonist which functions by alleviating the effects of exorbitant amounts of glutamate: a chemical found in the brain (Mathew et al., 2019). Furthermore, there was a targeted therapy approved for treating AD is a disease-modifying medication such as aducanumab: an A β -directed antibody (Thomas, Wasunna-Smith, & Kuruvilla, 2021). Aducanumab targets the A β protein in the brain of AD patients, causing the reduction of amyloid plaques. Currently, there are ongoing studies as to whether this medication is suitable and effective on patients as they progress through the stages of cognitive impairment (Thomas et al., 2021). Therefore, attempts to develop novel strategies for AD treatment such as stem cell therapy are ongoing for finding effective therapies for AD (Thomas et al., 2021).

Stem cell classes based on cell origin

There are four classes of stem cells that are frequently employed in current AD experimentation and studies: (1) embryonic stem cells (ESCs), (2) Mesenchymal stem cells (MSCs), (3) Brain-derived neural stem cells (NSCs), and (4) induced pluripotent stem cells (iPSCs) (S. U. Kim, Lee, & Kim, 2013; Q. Liu et al., 2012; Titomanlio et al., 2011). ESCs are acquired from the inner cells of the blastocyst and are classified as pluripotent stem cells due to their capacity to produce different cell types from the germ layers. iPSCs are commonly obtained from mature adult dermal fibroblasts, in vitro, by laboratory processes (Cho et al., 2010; Hu et al., 2020; Stadtfeld et al., 2010). The genetic alteration of these stem cells provides them with pluripotent capabilities and ESC-like phenotypes and differentiation capacities. MSCs can be derived from a variety of sources, starting with umbilical cord blood (ucb-MSCs), the Wharton jelly, and adult stem cell pupae such as bone marrow and adipose tissue (Zeng et al., 2013). Research done using rodent models of AD and ucb-MSCs have found that the transplantation of ucb-MSCs improved spatial learning and had prevented memory decline in the murine models. The research further proposed additional mechanisms of the transplantation such as the reduction of A β plaques, hyperphosphorylation of tau protein, and microglial inflammation, as well as the increase of anti-inflammatory cytokines (Cho et al., 2010; Hei et al., 2017). MSCs have been recorded to have immunomodulatory and anti-inflammatory effects through their ability to upregulate neuroprotection and downregulate pro-inflammatory cytokines (Guo et al., 2020; Prasad, 2017; Zeng et al., 2013).

Mesenchymal stem cell therapeutic strategy in Alzheimer's Disease

MSCs are obtained through bone marrow, adipose tissue, and the tissue of the umbilical cord; its alleged mechanism and prerequisites include (1) homing, (2) paracrine neuroprotective agents, and (3) its immunoregulation (Jin et al., 2013; Kern, Eichler, Stoeve, Klüter, & Bieback, 2006; Stanko, Kaiserova, Altanerova, & Altaner, 2014). Though the mechanisms of MSCs are currently still being researched, in light of recent studies, these therapeutic mechanisms proved to have the potential to ameliorate various diseases in innumerable ways

(Oh, Son, Choi, Lee, & Byun, 2018). In the present, there has been a limited indication for both the function and neural maturation of MSC-derived neurons *in vivo*. Additionally, the neuroplacement by MSCs is left in its infancy due to low rates of neuronal differentiation (Donega et al., 2014; Tanna & Sachan, 2014; Wagenaar et al., 2018). Though these aspects cause uncertainty and hesitance in the development of MSC-based therapy, studies of MSCs show potential for the future of neuroregenerative treatments (Donega et al., 2014; Tanna & Sachan, 2014; Wagenaar et al., 2018).

MSCs have attested to improving the pathological factors characterizing AD. In a previous study, A β was injected into the dentate gyrus of mice, inducing a model of acute AD. Bone marrow-derived mesenchymal stem cells (BM-MSCs) were also intracerebrally transplanted into the affected area causing the activation of microglia near the site. This sprung the diminution of A β deposits and enhanced cognitive function, demonstrating the potential of BM-MSCs as a therapeutic agent against AD (Bae, Jin, Lee, Richardson, & Carter, 2013; Neves et al., 2021; Tanna & Sachan, 2014). Human placenta amniotic membrane-derived mesenchymal stem cells (hAMMSCs) were injected intravenously into AD model mice, causing significantly improved spatial learning and function as well as the reduction of amyloid plaques in the brain (Kharat et al., 2019; Neves et al., 2021; Tanna & Sachan, 2014). This was shown through the decreased levels of oxidative stress, indicating a correlation between the improved symptoms of AD and the regulation of oxidative stress (Jiao et al., 2016). Several studies have illustrated the potential of hAMMSCs treatment on the immunology of AD and hippocampal function through oxidative stress (Jiao et al., 2016; Shariati et al., 2020). Intracerebral transplantation of adipose-derived mesenchymal stem cells (ADSCs) poses as a propitious cell genesis for regenerative therapy (Mazini, Rochette, Amine, & Malka, 2019). Results from a research have shown that usage of ADSC transplantation has drastically reduced A β peptide deposition along with restoration of cognitive function lost from AD (Jiao et al., 2016; Ma et al., 2013; Mazini et al., 2019; Shariati et al., 2020; Yan et al., 2014). Research done on human adipose tissue-derived mesenchymal stem cells (ADMSCs) being either intravenously or intracerebroventricularly

injected into the 18-month-old mice via the brain; this resulted in improved locomotor activity and neurological function in the animals. A study with respect to the transplantation of MSCs into aged rodent models showed that the cells underwent differentiation into various types of neural cells (Jiao et al., 2016; Shariati et al., 2020). With increased levels of neural cells, this resulted in increased concentrations of ACh neurotransmitter, BDNF, and NGF; thus, bringing about improved locomotor and cognitive function in the rodents (Gericke, 2020; Latham, Wang, Patterson, Slikker Jr, & Liu, 2021; Y.-X. Li, Ye, Chen, Jia, & He, 2018; Popova, Kulikov, & Naumenko, 2020).

MSCs can offer therapeutic roles in the design of cell replacement. Chem-Cun Wu et al. report that BM-MSCs -induced by a granulocyte colony-stimulating factor- were able to replace the neural lineage cells (Wu et al., 2017). This discovery exhibits tremendous potential in MSC treatment for the restoration of neurological function in mice models of AD (Wu et al., 2017). Furthering the potential of MSCs in neurodegenerative treatment; experimentation in the topic regarding the role of cytokines and signaling pathways in MSCs has gained substantial attention in the past years. Research conducted by Raheleh Farahzadi et al. have found when BM-MSCs have been co-cultured with A β -treated primary cortical and hippocampal neural cells (as an *in vitro* cell line model for AD), there was observed increase of cytokine secretion and telomere length (Farahzadi, Fathi, & Vietor, 2020). These two factors were regarded as a hallmark of neurodegenerative disorder as well as telomerase activity (Farahzadi et al., 2020).

Stem cell trials in humans

The fact that there have been discrepancies in preclinical research have limited some prospective stem cell therapies from advancing to human clinical trials is a barrier to the success of those therapies (Duncan & Valenzuela, 2017; Klionsky et al., 2021; X.-Y. Liu, Yang, & Zhao, 2020). In contrast, evidence from animal model studies, as well as the simplicity of handling and isolation, has contributed to human clinical trials approving MSC-based therapies (Duncan & Valenzuela, 2017). A recent open-label phase I clinical trial (involving patients who received intravenous injections of human umbilical cord blood-derived mesenchymal stem

cells (hUCB-MSCs)) assessed the welfare and tolerability of intracranially injecting allogeneic hUCB-MSCs (Trial identifiers: NCT01297218 and NCT01696591) (Duncan & Valenzuela, 2017). NINDS-ADRA criteria were used to classify nine people with probable AD as participants in the trial (Nicolaou *et al.*, 2001). The following criteria were used to determine who should participate in the study: 10 and 24 score in the Mini-Mental State Examination which indicates mild-moderate AD dementia, along with the Pittsburgh compound B positron emission tomography analysis to verify the presence of amyloid immunology. After receiving an injection of three million cells into the hippocampus and precuneus, participants were separated into a low-dose group (about three million cells) and a high-dose group (approximately six million cells) (Duncan & Valenzuela, 2017). After 3 months and 24 months, none of the patients experienced significant side effects, such as complications from the surgical surgery or MSC treatment (Duncan & Valenzuela, 2017). MSC transplantation, however, failed to impede neurological deterioration throughout the 2-year investigation, as indicated by the Alzheimer's Disease Assessment Scale-cognitive subscale, therefore, it lacked any negative effect on the population. Additional alterations to AD pathology were found not to have occurred. This means that the immunomodulatory impact of MSCs, commonly described in animal models of AD [30–32], was not apparent (Duncan & Valenzuela, 2017). They believe this may partly be due to dependence on neuroimaging, which is more difficult to interpret in live subjects, instead of more sensitive post-mortem biochemical analysis utilised in animal research (Duncan & Valenzuela, 2017).

MSCs produced from the umbilical cord blood are still commonly utilised. However, there are notable differences in the number of cells, and the delivery schedule. Two distinct trials are now running and recruiting, each utilising MSC sources that are different from each other. In a study of the welfare and effectiveness of adipose-derived stromal vascular fraction cells collected following liposuction procedures, one trial (Trial identifier: NCT02912169) will use participants who have had such procedures. Ischaemia-tolerant allogeneic human bone marrow-derived MSCs will be the subjects of a new study (Trial identifier: NCT02833792) (Duncan &

Valenzuela, 2017). To be more like the physiological milieu of the central nervous system, these MSCs were grown in hypoxic circumstances, which increase the expression of angiogenic growth factors such as VEGF and angiopoietin. As a result, these cells demonstrate improved migratory behaviour (Duncan & Valenzuela, 2017).

Future directions

Preclinical research implies that stem cells can cure AD; nevertheless, the difficulty in translation between animal and human studies holds back the use of stem cells in this area. Several research groups have found numerous approaches to treat AD in transgenic mouse models. Regrettably, all of these techniques have disappointed in human clinical trials (Duncan & Valenzuela, 2017). While genetically engineered models are far from being valuable predictors, they provide little if any predictive utility (Duncan & Valenzuela, 2017). The majority of human AD arises in a markedly heterogeneous population, while genetically engineered models are mainly based on family AD-related assumptions in a genetically homogeneous population (Duncan & Valenzuela, 2017; Klionsky *et al.*, 2021). At this point, the use of rat models and associated theoretical causal hypotheses for human diseases is incompetent for predicting human clinical outcomes. Additionally, the neurons and synapses they use to simulate AD fail to reiterate the loss of neuronal function that is crucial to AD. Because AD cell therapies will be required to illustrate success in animals which mirrors the scientific and neurodegenerative aspects of human AD, they will be required to demonstrate clinical and neurodegenerative AD-like symptoms in additional animals (Yan *et al.*, 2014).

Other critical problems must also be answered, including long-term safety, the best source of cells, and the delivery technology. The pathogenic AD environment must also be evaluated and understood. In the investigations covered below, heterotopic stem cells were employed. Transplantation of allogeneic foetal NSC, genetically modified MSC, and autologous haematopoietic stem cells has led to tumour growth in rodents and humans. It is also clinically relevant since NSC niches remain inaccessible in adults (Duncan & Valenzuela, 2017). While replacing lost neural circuits with neuron reconstruction therapies may not restore lost neural

function, it can temporarily augment existing depleted circuits enough to help improve cognitive performance, everyday functioning, and overall quality of life (Farahzadi et al., 2020). Individuals with AD dementia have a lifespan of approximately four to five years, which translates to a functional cure if a near replacement therapy can rescue and protect brain function over this time. Even though many pathophysiological factors contribute to the AD process, an integrated treatment approach could be required, using pharmacological approaches to address pathology, endogenous neurogenesis and synaptogenesis, and the administration of exogenous neuro resources substances (Duncan & Valenzuela, 2017; X.-Y. Liu et al., 2020).

Conclusion

Stem cell therapy for Alzheimer's disease holds significant potential, although it is still in the early stages of development. New investigations continue to discover treatment mechanisms as new preclinical research has substantiated proof-of-concept. Most clinical trials to date, when employing MSC-based therapies, have been relatively consistent. At present, one such trial has failed, but many others are ongoing. It is essential to understand, however, that there is an enormous chasm between rats and humans. However, before this project gets off the ground, we will need to acknowledge the cells and the brains they want to heal and use models that begin to bridge this gap.

References

1. Aanandhi, M. Vijey, Niventhi, A., Rujaswini, T., Hemalatha, C. N., & Praveen, D. (2018). A Comprehensive Review on the Role of Tau Proteins in Alzheimer's Pathology. *Research Journal of Pharmacy and Technology*, 11(2), 788-790.
2. Alavi Naini, Seyedeh Maryam, & Soussi-Yanicostas, Nadia. (2015). Tau hyperphosphorylation and oxidative stress, a critical vicious circle in neurodegenerative tauopathies? *Oxidative medicine and cellular longevity*, 2015.
3. Azevedo, E. P., Ledo, J. H., Barbosa, G., Sobrinho, M., Diniz, L., Fonseca, A. C. C., . . . Palhano, F. L. (2013). Activated microglia mediate synapse loss and short-term memory deficits in a mouse model of transthyretin-related oculoleptomeningeal amyloidosis. *Cell death & disease*, 4(9), e789-e789.
4. Baas, Peter W., & Qiang, Liang. (2019). Tau: it's not what you think. *Trends in cell biology*, 29(6), 452-461.
5. Bae, Jae-sung, Jin, Hee Kyung, Lee, Jong Kil, Richardson, Jill C., & Carter, Janet E. (2013). Bone marrow-derived mesenchymal stem cells contribute to the reduction of amyloid- β deposits and the improvement of synaptic transmission in a mouse model of pre-dementia Alzheimer's disease. *Current Alzheimer Research*, 10(5), 524-531.
6. Bayer, Thomas A., & Wirths, Oliver. (2010). Intracellular accumulation of amyloid-Beta-a predictor for synaptic dysfunction and neuron loss in Alzheimer's disease. *Frontiers in aging neuroscience*, 2, 8.
7. Bessis, Alain, Béchade, Catherine, Bernard, Delphine, & Roumier, Anne. (2007). Microglial control of neuronal death and synaptic properties. *Glia*, 55(3), 233-238.
8. Bhandare, Vishwambhar Vishnu, Kumbhar, Bajarang Vasant, & Kunwar, Ambarish. (2019). Differential binding affinity of tau repeat region R2 with neuronal-specific β -tubulin isoforms. *Scientific reports*, 9(1), 1-12.
9. Bondi, Mark W., Edmonds, Emily C., & Salmon, David P. (2017). Alzheimer's disease: past, present, and future. *Journal of the International Neuropsychological Society*, 23(9-10), 818-831.
10. Boonen, R. A., van Tijn, P., & Zivkovic, D. (2009). Wnt signaling in Alzheimer's disease: up or down, that is the question. *Ageing Res Rev*, 8(2), 71-82. doi:10.1016/j.arr.2008.11.003
11. Bossenmeyer-Pouricé, Carine, Smith, A. David, Lehmann, Sylvain, Deramecourt, Vincent, Sablonnière, Bernard, Camadro, Jean-Michel, . . . Umoret, Remy. (2019). N-homocysteinylation of tau and MAP1 is increased in autopsy specimens of Alzheimer's disease and vascular dementia. *The Journal of pathology*, 248(3), 291-303.
12. Briggs, Robert, Kennelly, Sean P., & O'Neill, Desmond. (2016). Drug treatments in Alzheimer's disease. *Clinical medicine*, 16(3), 247.
13. Chang, Keun- A., Kim, Hee Jin, Joo, Yuyoung, Ha, Sungji, & Suh, Yoo-Hun. (2014). The

- therapeutic effects of human adipose-derived stem cells in Alzheimer's disease mouse models. *Neurodegenerative Diseases*, 13(2-3), 99-102.
14. Cho, Hyun-Jai, Lee, Choon-Soo, Kwon, Yoo-Wook, Paek, Jae Seung, Lee, Sun-Hee, Hur, Jin, . . . Leem, Sun-Hee. (2010). Induction of pluripotent stem cells from adult somatic cells by protein-based reprogramming without genetic manipulation. *Blood, The Journal of the American Society of Hematology*, 116(3), 386-395.
 15. Chong, Fong Ping, Ng, Khuen Yen, Koh, Rhun Yian, & Chye, Soi Moi. (2018). Tau proteins and tauopathies in Alzheimer's disease. *Cellular and molecular neurobiology*, 38(5), 965-980.
 16. Connors, Michael H., Ames, David, Woodward, Michael, & Brodaty, Henry. (2018). Psychosis and clinical outcomes in Alzheimer disease: a longitudinal study. *The American Journal of Geriatric Psychiatry*, 26(3), 304-313.
 17. Deng, Shengqiong, Ge, Zhiru, Song, Yuting, Wang, Hairong, Liu, Xinghui, & Zhang, Denghai. (2019). Exosomes from adipose-derived mesenchymal stem cells ameliorate cardiac damage after myocardial infarction by activating S1P/SK1/S1PR1 signaling and promoting macrophage M2 polarization. *The international journal of biochemistry & cell biology*, 114, 105564.
 18. Donega, Vanessa, Nijboer, Cora H., Braccioli, Luca, Slaper-Cortenbach, Ineke, Kavelaars, Annemieke, Van Bel, Frank, & Heijnen, Cobi J. (2014). Intranasal administration of human MSC for ischemic brain injury in the mouse: in vitro and in vivo neuroregenerative functions. *PLoS one*, 9(11), e112339.
 19. Drepper, Friedel, Biernat, Jacek, Kaniyappan, Senthilvelrajan, Meyer, Helmut E., Mandelkow, Eva Maria, Warscheid, Bettina, & Mandelkow, Eckhard. (2020). A combinatorial native MS and LC-MS/MS approach reveals high intrinsic phosphorylation of human Tau but minimal levels of other key modifications. *Journal of Biological Chemistry*, 295(52), 18213-18225.
 20. Duncan, Thomas, & Valenzuela, Michael. (2017). Alzheimer's disease, dementia, and stem cell therapy. *Stem cell research & therapy*, 8(1), 1-9.
 21. Esmieu, Charlène, Ferrand, Guillaume, Borghesani, Valentina, & Hureau, Christelle. (2021). Impact of N-Truncated A β Peptides on Cu-and Cu (A β)-Generated ROS: CuI Matters! *Chemistry—A European Journal*, 27(5), 1777-1786.
 22. Farahzadi, Raheleh, Fathi, Ezzatollah, & Vietor, Ilja. (2020). Mesenchymal stem cells could be considered as a candidate for further studies in cell-based therapy of Alzheimer's disease via targeting the signaling pathways. *ACS chemical neuroscience*, 11(10), 1424-1435.
 23. Feng, Qiong, Luo, Yu, Zhang, Xiang-Nan, Yang, Xi-Fei, Hong, Xiao-Yue, Sun, Dong-Shen, . . . Zhang, Jun-Fei. (2020). MAPT/Tau accumulation represses autophagy flux by disrupting IST1-regulated ESCRT-III complex formation: a vicious cycle in Alzheimer neurodegeneration. *Autophagy*, 16(4), 641-658.
 24. Fontana, Igor C., Zimmer, Aline R., Rocha, Andreia S., Gosmann, Grace, Souza, Diogo O., Lourenco, Mychael V., . . . Zimmer, Eduardo R. (2020). Amyloid- β oligomers in cellular models of Alzheimer's disease. *Journal of neurochemistry*, 155(4), 348-369.
 25. Gant, John C., Kadish, Inga, Chen, Kuey-Chu, Thibault, Olivier, Blalock, Eric M., Porter, Nada M., & Landfield, Philip W. (2018). Aging-related calcium dysregulation in rat entorhinal neurons homologous with the human entorhinal neurons in which Alzheimer's disease neurofibrillary tangles first appear. *Journal of Alzheimer's Disease*, 66(4), 1371-1378.
 26. Gericke, Johané. (2020). Evaluating the antidepressant-like properties of *Sceletium tortuosum*, alone and as adjunctive treatment.
 27. Guo, Min-fang, Zhang, Hui-yu, Li, Yan-hua, Gu, Qing-fang, Wei, Wen-yue, Wang, Yu-yin, . . . Chai, Zhi. (2020). Fasudil inhibits the activation of microglia and astrocytes of transgenic Alzheimer's disease mice via the downregulation of TLR4/Myd88/NF- κ B pathway. *Journal of Neuroimmunology*, 346, 577284.
 28. Hanfelt, John J., Peng, Limin, Goldstein, Felicia C., & Lah, James J. (2018). Latent classes of mild cognitive impairment are associated with clinical outcomes and neuropathology: Analysis of data from the National Alzheimer's

- Coordinating Center. *Neurobiology of disease*, 117, 62-71.
29. Hanger, Diane P., Betts, Joanna C., Loviny, Th  r  se L. F., Blackstock, Walter P., & Anderton, Brian H. (1998). New phosphorylation sites identified in hyperphosphorylated tau (paired helical filament-tau) from Alzheimer's disease brain using nanoelectrospray mass spectrometry. *Journal of neurochemistry*, 71(6), 2465-2476.
 30. Hefti, Marco M., Kim, SoongHo, Bell, Aaron J., Betters, Ryan K., Fiock, Kimberly L., Iida, Megan A., . . . Crary, John F. (2019). Tau phosphorylation and aggregation in the developing human brain. *Journal of Neuropathology & Experimental Neurology*, 78(10), 930-938.
 31. Hei, Wei-hong, Almansoori, Akram A., Sung, Mi-Ae, Ju, Kyung-Won, Seo, Nari, Lee, Sung-Ho, . . . He, Hong. (2017). Adenovirus vector-mediated ex vivo gene transfer of brain-derived neurotrophic factor (BDNF) to human umbilical cord blood-derived mesenchymal stem cells (UCB-MSCs) promotes crush-injured rat sciatic nerve regeneration. *Neuroscience letters*, 643, 111-120.
 32. Hu, Zhixing, Li, Hanqin, Jiang, Houbo, Ren, Yong, Yu, Xinyang, Qiu, Jingxin, . . . Feng, Jian. (2020). Transient inhibition of mTOR in human pluripotent stem cells enables robust formation of mouse-human chimeric embryos. *Science Advances*, 6(20), eaaz0298.
 33. Ji, Minbiao, Arbel, Michal, Zhang, Lili, Freudiger, Christian W., Hou, Steven S., Lin, Dongdong, . . . Xie, X. Sunney. (2018). Label-free imaging of amyloid plaques in Alzheimer's disease with stimulated Raman scattering microscopy. *Science advances*, 4(11), eaat7715.
 34. Jiang, Conglin, Zou, Xiang, Zhu, Renqing, Shi, Yimin, Wu, Zehan, Zhao, Fan, & Chen, Liang. (2018). The correlation between accumulation of amyloid beta with enhanced neuroinflammation and cognitive impairment after intraventricular hemorrhage. *Journal of neurosurgery*, 131(1), 54-63.
 35. Jiao, Hongliang, Shi, Ke, Zhang, Weijie, Yang, Liang, Yang, Lu, Guan, Fangxia, & Yang, Bo. (2016). Therapeutic potential of human amniotic membrane-derived mesenchymal stem cells in APP transgenic mice. *Oncology letters*, 12(3), 1877-1883.
 36. Jin, Hye Jin, Bae, Yun Kyung, Kim, Miyeon, Kwon, Soon-Jae, Jeon, Hong Bae, Choi, Soo Jin, . . . Chang, Jong Wook. (2013). Comparative analysis of human mesenchymal stem cells from bone marrow, adipose tissue, and umbilical cord blood as sources of cell therapy. *International journal of molecular sciences*, 14(9), 17986-18001.
 37. Kellogg, Elizabeth H., Hejab, Nisreen M. A., Poepsel, Simon, Downing, Kenneth H., DiMaio, Frank, & Nogales, Eva. (2018). Near-atomic model of microtubule-tau interactions. *Science*, 360(6394), 1242-1246.
 38. Kern, Susanne, Eichler, Hermann, Stoeve, Johannes, Kl  ter, Harald, & Bieback, Karen. (2006). Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. *Stem cells*, 24(5), 1294-1301.
 39. Kharat, Avinash, Chandravanshi, Bhawna, Gadre, Shashikant, Patil, Vikrant, Bhonde, Ramesh, & Dubhashi, Aparna. (2019). IGF-1 and somatocinin trigger islet differentiation in human amniotic membrane derived mesenchymal stem cells. *Life sciences*, 216, 287-294.
 40. Kim, Seung U., Lee, Hong J., & Kim, Yun B. (2013). Neural stem cell-based treatment for neurodegenerative diseases. *Neuropathology*, 33(5), 491-504.
 41. Kim, Yoo Sung, Jung, Hae Myeong, & Yoon, Bo-Eun. (2018). Exploring glia to better understand Alzheimer's disease. *Animal cells and systems*, 22(4), 213-218.
 42. Kimura, Tetsuya, Whitcomb, Daniel J., Jo, Jihoon, Regan, Philip, Piers, Thomas, Heo, Seonghoo, . . . Seok, Heon. (2014). Microtubule-associated protein tau is essential for long-term depression in the hippocampus. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 369(1633), 20130144.
 43. Klionsky, D. J., Abdel-Aziz, A. K., Abdelfatah, S., Abdellatif, M., Abdoli, A., Abel, S., . . . Tong, C. K. (2021). Guidelines for the use and interpretation of assays for monitoring autophagy (4th edition). *Autophagy*, 17(1), 1-382. doi:10.1080/15548627.2020.1797280

44. Latham, Leah E., Wang, Cheng, Patterson, Tucker A., Slikker Jr, William, & Liu, Fang. (2021). Neuroprotective Effects of Carnitine and Its Potential Application to Ameliorate Neurotoxicity. *Chemical Research in Toxicology*.
45. Li, Shaomin, & Selkoe, Dennis J. (2020). A mechanistic hypothesis for the impairment of synaptic plasticity by soluble A β oligomers from Alzheimer's brain. *Journal of neurochemistry*, 154(6), 583-597.
46. Li, Yi-Xuan, Ye, Zi-He, Chen, Tong, Jia, Xiao-Feng, & He, Ling. (2018). The effects of donepezil on phencyclidine-induced cognitive deficits in a mouse model of schizophrenia. *Pharmacology Biochemistry and Behavior*, 175, 69-76.
47. Liu, Qiuyue, Spusta, Steven C., Mi, Ruifa, Lassiter, Rhonda N. T., Stark, Michael R., Höke, Ahmet, . . . Zeng, Xianmin. (2012). Human neural crest stem cells derived from human ESCs and induced pluripotent stem cells: induction, maintenance, and differentiation into functional schwann cells. *Stem cells translational medicine*, 1(4), 266-278.
48. Liu, Xin-Yu, Yang, Lin-Po, & Zhao, Lan. (2020). Stem cell therapy for Alzheimer's disease. *World Journal of Stem Cells*, 12(8), 787.
49. Long, Justin M., & Holtzman, David M. (2019). Alzheimer disease: an update on pathobiology and treatment strategies. *Cell*, 179(2), 312-339.
50. Ma, Tuo, Gong, Kai, Ao, Qiang, Yan, Yufang, Song, Bo, Huang, Hongyun, . . . Gong, Yandao. (2013). Intracerebral transplantation of adipose-derived mesenchymal stem cells alternatively activates microglia and ameliorates neuropathological deficits in Alzheimer's disease mice. *Cell transplantation*, 22(1_suppl), 113-126.
51. Magalhães, Paulo, Alves, Gilberto, Fortuna, Ana, Llerena, Adrián, & Falcão, Amílcar. (2020). Real-world clinical characterization of subjects with depression treated with antidepressant drugs focused on (non-) genetic factors, pharmacokinetics, and clinical outcomes: GnG-PK/PD-AD study. *Experimental and clinical psychopharmacology*, 28(2), 202.
52. Martins, Antonio Henrique, Zayas-Santiago, Astrid, Ferrer-Acosta, Yancy, Martinez-Jimenez, Solianne M., Zueva, Lidia, Diaz-Garcia, Amanda, & Inyushin, Mikhail. (2019). Accumulation of amyloid beta (A β) peptide on blood vessel walls in the damaged brain after transient middle cerebral artery occlusion. *Biomolecules*, 9(8), 350.
53. Mathew, Bijo, Parambi, Della G. T., Mathew, Githa E., Uddin, Md Sahab, Inasu, Sini T., Kim, Hoon, . . . Carradori, Simone. (2019). Emerging therapeutic potentials of dual-acting MAO and AChE inhibitors in Alzheimer's and Parkinson's diseases. *Archiv der Pharmazie*, 352(11), 1900177.
54. Mazini, Loubna, Rochette, Luc, Amine, Mohamed, & Malka, Gabriel. (2019). Regenerative capacity of adipose derived stem cells (ADSCs), comparison with mesenchymal stem cells (MSCs). *International journal of molecular sciences*, 20(10), 2523.
55. Morris, Sarah L., Tsai, Ming-Ying, Aloe, Sarah, Bechberger, Karin, König, Svenja, Morfini, Gerardo, & Brady, Scott T. (2021). Defined tau phosphospecies differentially inhibit fast axonal transport through activation of two independent signaling pathways. *Frontiers in Molecular Neuroscience*, 13, 269.
56. Murray, Ana P., Faraoni, María Belén, Castro, María Julia, Alza, Natalia Paola, & Cavallaro, Valeria. (2013). Natural AChE inhibitors from plants and their contribution to Alzheimer's disease therapy. *Current neuropharmacology*, 11(4), 388-413.
57. Nam, Eunjoo, Lee, Yeong-Bae, Moon, Cheil, & Chang, Keun- A. (2020). Serum tau proteins as potential biomarkers for the assessment of Alzheimer's disease progression. *International journal of molecular sciences*, 21(14), 5007.
58. Narayanan, S. E., Sekhar, N., Rajamma, R. G., Marathakam, A., Al Mamun, A., Uddin, M. S., & Mathew, B. (2020). Exploring the Role of Aggregated Proteomes in the Pathogenesis of Alzheimer's Disease. *Curr Protein Pept Sci*, 21(12), 1164-1173. doi:10.2174/1389203721666200921152246
59. Nasu, Ryuto, Furukawa, Ayako, Suzuki, Keita, Takeuchi, Masayoshi, & Koriyama, Yoshiki. (2020). The Effect of Glyceraldehyde-Derived Advanced Glycation End Products on β -Tubulin-Inhibited Neurite Outgrowth in SH-SY5Y

- Human Neuroblastoma Cells. *Nutrients*, 12(10), 2958.
60. Neves, Amanda Ferreira, Camargo, Christian, Premer, Courtney, Hare, Joshua M., Baumel, Bernard S., & Pinto, Milena. (2021). Intravenous administration of mesenchymal stem cells reduces Tau phosphorylation and inflammation in the 3xTg-AD mouse model of Alzheimer's disease. *Experimental neurology*, 341, 113706.
 61. Nicolaou, M., Song, Y. Q., Sato, C., Orlacchio, A., Kawarai, T., Medeiros, Helena, . . . Rogaeve, E. (2001). Mutations in the open reading frame of the β -site APP cleaving enzyme (BACE) locus are not a common cause of Alzheimer's disease. *Neurogenetics*, 3(4), 203-206.
 62. Nu, Truong Thi Vu, Tran, Nhu Hoa Thi, Nam, Eunjoon, Nguyen, Tan Tai, Yoon, Won Jung, Cho, Sungbo, . . . Ju, Heongkyu. (2018). Blood-based immunoassay of tau proteins for early diagnosis of Alzheimer's disease using surface plasmon resonance fiber sensors. *RSC advances*, 8(14), 7855-7862.
 63. Oh, Seyeon, Son, Myeongjoo, Choi, Junwon, Lee, Sojung, & Byun, Kyunghye. (2018). sRAGE prolonged stem cell survival and suppressed RAGE-related inflammatory cell and T lymphocyte accumulations in an Alzheimer's disease model. *Biochemical and biophysical research communications*, 495(1), 807-813.
 64. Pascoal, Tharick A., Shin, Monica, Kang, Min Su, Chamoun, Mira, Chartrand, Daniel, Mathotaarachchi, Sulantha, . . . Hopewell, Robert. (2018). In vivo quantification of neurofibrillary tangles with [18 F] MK-6240. *Alzheimer's research & therapy*, 10(1), 1-14.
 65. Pascoal, Tharick A., Therriault, Joseph, Benedet, Andrea L., Savard, Melissa, Lussier, Firoza Z., Chamoun, Mira, . . . Mathotaarachchi, Sulantha. (2020). 18F-MK-6240 PET for early and late detection of neurofibrillary tangles. *Brain*, 143(9), 2818-2830.
 66. Pluta, Ryszard, Furmaga-Jabłońska, Wanda, Maciejewski, Ryszard, Ułamek-Kozioł, Marzena, & Jabłoński, Mirosław. (2013). Brain ischemia activates β - and γ -secretase cleavage of amyloid precursor protein: significance in sporadic Alzheimer's disease. *Molecular neurobiology*, 47(1), 425-434.
 67. Popova, Nina K., Kulikov, Alexander V., & Naumenko, Vladimir S. (2020). Spaceflight and brain plasticity: Spaceflight effects on regional expression of neurotransmitter systems and neurotrophic factors encoding genes. *Neuroscience & Biobehavioral Reviews*.
 68. Prasad, Kedar N. (2017). Oxidative stress and pro-inflammatory cytokines may act as one of the signals for regulating microRNAs expression in Alzheimer's disease. *Mechanisms of ageing and development*, 162, 63-71.
 69. Qiang, Liang, Sun, Xiaohuan, Austin, Timothy O., Muralidharan, Hemalatha, Jean, Daphney C., Liu, Mei, . . . Baas, Peter W. (2018). Tau does not stabilize axonal microtubules but rather enables them to have long labile domains. *Current Biology*, 28(13), 2181-2189.
 70. Rana, Monika, & Sharma, Anuj Kumar. (2019). Cu and Zn interactions with A β peptides: consequence of coordination on aggregation and formation of neurotoxic soluble A β oligomers. *Metallomics*, 11(1), 64-84.
 71. Samra, Elias Bou, Buhagiar-Labarchède, Géraldine, Machon, Christelle, Guitton, Jérôme, Onclercq-Delic, Rosine, Green, Michael R., . . . Amor-Guéret, Mounira. (2017). A role for Tau protein in maintaining ribosomal DNA stability and cytidine deaminase-deficient cell survival. *Nature communications*, 8(1), 1-14.
 72. Schauenburg, Linda, Liebsch, Filip, Eravci, Murat, Mayer, Magnus C., Weise, Christoph, & Multhaup, Gerhard. (2018). APLP1 is endoproteolytically cleaved by γ -secretase without previous ectodomain shedding. *Scientific reports*, 8(1), 1-12.
 73. Shariati, Ali, Nemati, Reza, Sadeghipour, Yasin, Yaghoubi, Yoda, Baghbani, Reza, Javidi, Kamran, . . . Hassanzadeh, Ali. (2020). Mesenchymal stromal cells (MSCs) for neurodegenerative disease: A promising frontier. *European journal of cell biology*, 99(6), 151097.
 74. Shigematsu, Kazuo, Takeda, Takahisa, Komori, Naoyuki, Tahara, Kenichi, & Yamagishi, Hisakazu. (2021). Hypothesis: Intravenous administration of mesenchymal stem cells is effective in the treatment of Alzheimer's disease. *Medical Hypotheses*, 150, 110572.
 75. Song, Ji-Won, & Choi, Byung-Sun. (2013). Mercury induced the accumulation of amyloid

- beta (A β) in PC12 cells: the role of production and degradation of A β . *Toxicological research*, 29(4), 235-240.
76. Stadtfeld, Matthias, Apostolou, Effie, Akutsu, Hidenori, Fukuda, Atsushi, Follett, Patricia, Natesan, Sridaran, . . . Hochedlinger, Konrad. (2010). Aberrant silencing of imprinted genes on chromosome 12qF1 in mouse induced pluripotent stem cells. *Nature*, 465(7295), 175-181.
 77. Stanko, Peter, Kaiserova, Katarina, Altanerova, Veronika, & Altaner, Cestmir. (2014). Comparison of human mesenchymal stem cells derived from dental pulp, bone marrow, adipose tissue, and umbilical cord tissue by gene expression. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub*, 158(3), 373-377.
 78. Tanna, Tanmay, & Sachan, Vatsal. (2014). Mesenchymal stem cells: potential in treatment of neurodegenerative diseases. *Current stem cell research & therapy*, 9(6), 513-521.
 79. Thomas, Emily, Wasunna-Smith, Brenda, & Kuruvilla, Tarun. (2021). Aducanumab and disease modifying treatments for Alzheimer's disease. *Progress in Neurology and Psychiatry*, 25(3), 4-6.
 80. Titomanlio, Luigi, Kavelaars, Annemieke, Dalous, Jeremie, Mani, Shyamala, El Ghouzzi, Vincent, Heijnen, Cobi, . . . Gressens, Pierre. (2011). Stem cell therapy for neonatal brain injury: perspectives and challenges. *Annals of neurology*, 70(5), 698-712.
 81. Vincent, Inez, Zheng, J. H., Dickson, D. W., Kress, Y., & Davies, P. (1998). Mitotic phosphoepitopes precede paired helical filaments in Alzheimer's disease. *Neurobiology of aging*, 19(4), 287-296.
 82. Wagenaar, Nienke, De Theije, Caroline G. M., De Vries, Linda S., Groenendaal, Floris, Benders, Manon J. N. L., & Nijboer, Cora H. A. (2018). Promoting neuroregeneration after perinatal arterial ischemic stroke: neurotrophic factors and mesenchymal stem cells. *Pediatric research*, 83(1), 372-384.
 83. Wang, Xin, & Pei, Gang. (2018). Visualization of Alzheimer's disease related α - β - γ -secretase ternary complex by bimolecular fluorescence complementation based fluorescence resonance energy transfer. *Frontiers in molecular neuroscience*, 11, 431.
 84. Weller, Jason, & Budson, Andrew. (2018). Current understanding of Alzheimer's disease diagnosis and treatment. *F1000Research*, 7.
 85. Wu, Cheng-Chun, Wang, I. Fang, Chiang, Po-Min, Wang, Liang-Chao, Shen, Che-Kun James, & Tsai, Kuen-Jer. (2017). G-CSF-mobilized bone marrow mesenchymal stem cells replenish neural lineages in Alzheimer's disease mice via CXCR4/SDF-1 chemotaxis. *Molecular neurobiology*, 54(8), 6198-6212.
 86. Yan, Yufang, Ma, Tuo, Gong, Kai, Ao, Qiang, Zhang, Xiufang, & Gong, Yandao. (2014). Adipose-derived mesenchymal stem cell transplantation promotes adult neurogenesis in the brains of Alzheimer's disease mice. *Neural Regeneration Research*, 9(8), 798.
 87. Zeng, Yu, Rong, Mingqiang, Liu, Yunsheng, Liu, Jingfang, Lu, Ming, Tao, Xiaoyu, . . . Li, Chuntao. (2013). Electrophysiological characterisation of human umbilical cord blood-derived mesenchymal stem cells induced by olfactory ensheathing cell-conditioned medium. *Neurochemical research*, 38(12), 2483-2489.
 88. Zhan, Xinhua, Stamova, Boryana, & Sharp, Frank R. (2018). Lipopolysaccharide associates with amyloid plaques, neurons and oligodendrocytes in Alzheimer's disease brain: a review. *Frontiers in aging neuroscience*, 10, 42